

Is Shorter Transient Middle Cerebral Artery Occlusion (t-MCAO) Duration Better in Stroke Experiments on Diabetic Female Sprague Dawely rats?

Mišir, Mihael; Renić, Marija; Mihalj, Martina; Novak, Sanja;
Drenjančević, Ines

Source / Izvornik: **Brain Injury, 2016, 30, 1390 - 6**

Journal article, Published version

Rad u časopisu, Objavljena verzija rada (izdavačev PDF)

<https://doi.org/10.1080/02699052.2016.1195518>

Permanent link / Trajna poveznica: <https://um.nsk.hr/um:nbn:hr:239:717761>

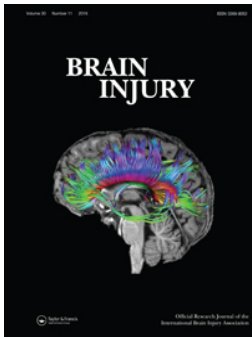
Rights / Prava: [In copyright](#)/[Zaštićeno autorskim pravom.](#)

Download date / Datum preuzimanja: **2025-01-22**



Repository / Repozitorij:

[Repository UHC Osijek - Repository University
Hospital Centre Osijek](#)



Is shorter transient middle cerebral artery occlusion (t-MCAO) duration better in stroke experiments on diabetic female Sprague Dawely rats?

Mihael Mišir, Marija Renić, Martina Mihalj, Sanja Novak & Ines Drenjančević

To cite this article: Mihael Mišir, Marija Renić, Martina Mihalj, Sanja Novak & Ines Drenjančević (2016) Is shorter transient middle cerebral artery occlusion (t-MCAO) duration better in stroke experiments on diabetic female Sprague Dawely rats?, *Brain Injury*, 30:11, 1390-1396, DOI: [10.1080/02699052.2016.1195518](https://doi.org/10.1080/02699052.2016.1195518)

To link to this article: <https://doi.org/10.1080/02699052.2016.1195518>



Published online: 19 Aug 2016.



Submit your article to this journal [↗](#)



Article views: 153



View related articles [↗](#)



View Crossmark data [↗](#)



Citing articles: 3 View citing articles [↗](#)

ORIGINAL ARTICLE

Is shorter transient middle cerebral artery occlusion (t-MCAO) duration better in stroke experiments on diabetic female Sprague Dawley rats?

Mihael Mišir^{1,2}, Marija Renić³, Martina Mihalj², Sanja Novak², & Ines Drenjančević²

¹Neurology Clinic, Clinical Hospital Center Osijek, Osijek, Croatia, ²Laboratory for Physiology of Circulation, Department of Physiology and Immunology, Faculty of Medicine, University Josip Juraj Strossmayer Osijek, Osijek, Croatia, and ³Croatian Institute for Brain Research, School of Medicine, University of Zagreb, Zagreb, Croatia

Abstract

Aim: To determine optimal duration of transient middle cerebral artery occlusion (t-MCAO) for a stroke model in female diabetic Sprague-Dawley (SD) rats.

Methods: Streptozotocin-induced type-1 diabetic SD female rats ($n = 25$, 12 weeks old, five groups; $n = 5$ per group) were subjected to different duration of t-MCAO (20, 30, 45, 60 and 90 minutes) followed by reperfusion. A control group of rats without diabetes ($n = 5$) was subjected to 30 minutes of t-MCAO followed by reperfusion. Twenty-four hours after reperfusion, infarct volumes were evaluated by 2,3,5-triphenyltetrazolium chloride (TTC) staining.

Results: Intra-ischaemic reductions of regional cerebral blood flow (rCBF) were similar in all groups (68–75% of baseline values). Reperfusion was significantly impaired in the 90-minute ischaemia group (56–62% vs 80–125% in other groups). Twenty minutes of t-MCAO induced a small infarct ($3 \pm 5\%$ of ischaemic hemisphere). Thirty minutes of t-MCAO produced a significantly larger infarct ($46 \pm 6\%$). In the 45 and 60 minute groups, ischaemia infarct was $52 \pm 5\%$ and $59 \pm 3\%$ of the ischaemic hemisphere, respectively. Ischaemia of 90' led to a massive stroke ($89 \pm 6\%$ of ischaemic hemisphere encompassing the whole striatum ($22 \pm 3\%$) and almost the whole MCA irrigated cortex area ($67 \pm 6\%$)). Thirty minutes of t-MCAO did not produce stroke in the control group.

Conclusion: The diabetic rat stroke model should be different from the non-diabetic, because female type-1 diabetic SD rats are highly sensitive to brain ischaemia and it is necessary to significantly shorten the duration of t-MCAO, optimally to 30 minutes.

Keywords

Model, cerebral ischaemia, transient, stroke, middle cerebral artery occlusion, diabetes, rats, female

History

Received 12 September 2015
Revised 4 February 2016
Accepted 7 May 2016
Published online 22 August 2016

Introduction

Approximately 25% of all stroke patients have diabetes mellitus and 40% have hyperglycaemia, which is associated with worse neurologic outcome as well as higher risk of recurrence of stroke [1,2]. Diabetic patients, compared to non-diabetics, are known to be more sensitive to cerebral ischaemia. Thus, the same duration of ischaemia results in more severe neurologic deficits and larger brain infarcts in diabetic patients. Female patients with diabetes mellitus have 4.8-fold higher risk for developing ischaemic stroke than the general population (compared to 3.7-fold for men) and more often suffer fatal strokes (standardized mortality ratios of 3.1 for males and 4.4 for females) [3–5]. The outcome is frequently lethal, regardless of any therapy undertaken, including recombinant tissue plasminogen activator (rtPA). Moreover, diabetes is in some cases, such as treatment of recurrent stroke with thrombolysis, one of exclusion criteria [6].

The most commonly used experimental model of stroke in rats is a model of middle cerebral artery occlusion

(MCAO) by intra-luminal suture. There are variations of this model in terms of use of permanent or transitory MCA occlusion-induced ischaemia. In rats without diabetes, duration of ischaemia in these models varies according to experiment from 60–120 minutes [7]. The duration of occlusion varies in models from permanent MCAO [8–10] to transitory MCAO (t-MCAO) of 180 [11], 120 [12–16], 105 [17] or 60 minutes [18]. Taking into account the observed differences in clinical presentation of diabetic vs non-diabetic patients with stroke, there are few issues that variations in experimental approach to stroke study are brought to light. For example, in diabetic rat stroke models, the same duration of MCAO as in non-diabetic rat models is used. Additionally, other models, e.g. the thromboembolic model, shows inconsistencies in the effects of occlusion which may be contributed to the unpredictable postponed reperfusion and consequently impossibility to predict with certainty the time when reperfusion will appear [19,20]. Such a model cannot simulate ischaemia in diabetic patients under internal carotid artery endarterectomy with accurately known duration of ischaemia and its localization in brain. On the other hand, the MCAO model with intra-luminal insertion of monofilament with LDF (laser Doppler flowmetry) monitoring in given time and known place,

Correspondence: Ines Drenjančević, MD, PhD, University Josip Juraj Strossmayer Osijek, Faculty of Medicine, Cara Hadrijana 10e, HR-31000 Osijek, Croatia. E-mail: ines.drenjancevic@mefos.hr

Color versions of one or more figures in the article can be found online at www.tandfonline.com/ibij.

mimics it perfectly and its complications alike [21,22]. With usual duration of t-MCAO used in non-diabetic rats, with all efforts and after significant time and animals used, it failed to establish stroke model in female rats with diabetes. Analysis found that the main reason for this was massive stroke with malignant brain oedema, producing devastating neurological deficits (such as inability to move, eat and drink) that became worse over time, leading to unconsciousness and death of animals within the first 24 hours. Taken all together, the aim of this study was to develop the adequate diabetic female rat model, using transitory middle cerebral artery occlusion (t-MCAO) that would produce treatable stroke conditions in rats with diabetes. To establish a reproducible model resulting with medium sized stroke, this study had to significantly shorten the duration of t-MCAO. The present study suggests that 30-minute t-MCAO could be a more appropriate stroke model than the usual 60–120 minute t-MCAO models.

Materials and methods

Animals

Experiments were performed on 30 Sprague-Dawley (SD) female rats, 12 weeks old, weighing between 220–272 g, housed in the accredited animal care facility at the University of Osijek. Animals were kept in cages with a 12-hour light/dark cycle and were allowed free access to food and water throughout the study. All experimental procedures were approved by the Ethical committee of the Faculty of Medicine, University of Osijek.

Induction of diabetes

Type-1 diabetes mellitus in rats was induced by a single intraperitoneal injection of streptozotocin (STZ; 60 mg kg⁻¹, Sigma-Aldrich, St. Louis, MO). STZ was freshly prepared in 0.09 M citrate saline buffer and administered to rats at 6 weeks of age [23,24]. Fasting blood glucose levels in rats were repeatedly measured using a glucometer (Accu-Check Active, Roche, Germany).

Study design

Diabetic rats that had fasting glucose levels above 17 mmol L⁻¹ were randomly assigned to five groups with different t-MCAO duration (20, 30, 45, 60 or 90 minutes) and non-diabetic rats were subjected to 30-minute t-MCAO ($n = 5$ animals per treatment group).

Exclusion criteria were: death of the animal during the surgical procedure or within 24 hours after reperfusion, excessive bleeding during operating procedures, reduction of rCBF during ischaemia less than 65% of baseline values, absence of reperfusion after removal of suture monitored using LDF, presence of intracerebral and/or subarachnoid haemorrhage at post-mortem examination of brains. Animals excluded from the study were replaced. Quantitative analysis of rCBF and infarct size was done by a researcher who was blinded to the experimental protocol.

Regional cerebral blood flow (rCBF) monitoring by laser Doppler flowmetry (LDF)

By medial approach the periosteum of parietal bones was exposed, hemostasis achieved and left parietal bone was thinned using a dental drill until translucent leaving the dura intact, 2 mm posterior and 6 mm lateral to the bregma. A laser Doppler flow probe was placed in a probe holder and fixed above the dura on the skull with glue and dental cement to continuously monitor cerebral blood flow in the cerebral cortex (LDF, Moor Instruments Ltd, model MBF3D, Devon, UK) during experiment.

Transient middle cerebral artery occlusion (t-MCAO)

Transient focal brain ischaemia was induced in the region of left MCA using a silicon-coated monofilament, as previously described by Koizumi et al. [25] and subsequently modified by Zea Longa et al. [26]. Anaesthesia was induced with intraperitoneal (i.p.) administration of combination of midazolam (Dormicum, Roche Pharma AG, Grenzach-Wyhlen, Germany) 0.5 mg kg⁻¹ and ketamine (Ketanest, PfizerPharma GmbH, Berlin, Germany) 75 mg kg⁻¹. Atropine (Atropini sulfas, Belupo d.d. Koprivnica, Croatia) 0.1 mg kg⁻¹ was also administered i.p. to reduce possible salivation as a side-effect of anaesthesia [27]. Animals were shaved, disinfected and fixed on the operating table and body temperature was continuously monitored with a rectal probe and maintained at $37 \pm 0.5^\circ\text{C}$ with a thermostatically regulated heating pad. After a midline incision of the neck was made, muscles were dislocated and common carotid artery (CCA), external carotid artery (ECA) and internal carotid artery (ICA) were carefully isolated from surrounding structures (nervus vagus and sympathetic plexus) under the operating microscope. CCA and ECA were ligated using 5-0 silk suture. A 4-0 nylon monofilament (coated at the tip with silicone 0.25–0.3 mm in diameter using silicone Xantopren and activator Elastomer, Heraeus Kulzer, Hanau, Germany) was introduced in the left ICA and gently advanced to the origin of the MCA until a slight resistance was felt and rCBF fell by ~ 70% of baseline value [28]. After a specified time of ischaemia (20–90 minutes) the filament was gently withdrawn to allow reperfusion. rCBF was continuously monitored to confirm adequate occlusion of the MCA and reperfusion after removal of the filament. After 20 minutes of reperfusion the incision on the neck was sutured, LDF probe removed and the animal was placed into the heated cage and monitored until recovered from anaesthesia. Animals had access to food and drink *ad libitum*.

Measurement of cerebral infarct volume

Twenty-four hours after reperfusion, the rats were deeply anaesthetized with ketamine and midazolam and killed by decapitation. The brains were quickly removed from the skull and placed into a steel brain matrix and were cut into 2 mm-thick coronal sections. The sections were stained with 2% 2,3,5-triphenyltetrazolium chloride (TTC, Sigma-Aldrich Chemie, Munich, Germany) at 37°C for 30 minutes in the dark. Viable brain tissue was stained dark brick red due to

reduction of TTC to formazan by mitochondrial enzymes, whereas necrotic brain areas were unstained and appeared white. The sections were fixed in 10% buffered formalin solution for 24 hours and anterior and posterior sides of each stained brain section were scanned. Infarct volumes were measured using Image J imaging software (v1.37, National Institute of Health, Bethesda, MD, USA) and expressed as a percentage of ischaemic hemisphere [29].

Statistical analysis

Data analysis was performed using SigmaPlot software (version 11.0, Systat Software Inc., San Jose, CA) and the graphs were created using GraphPad Prism (v5, GraphPad Software, Inc., La Jolla, CA). Shapiro-Wilks test was performed to assess normality of data distribution. Between-group analysis for normally distributed variables was tested using parametric one-way ANOVA and post-hoc Tukey test, whereas in the case of variables that violated assumption of normality differences were tested with non-parametric Kruskal–Wallis test followed by pairwise multiple comparison procedures (Dunn's and Student–Newman–Keuls method). Cerebral blood flow data were compared by two-way ANOVA and Bonferroni multiple comparison. Data are presented as mean and standard error of the mean (mean \pm SEM). Accepted statistical significance was for $p < 0.05$.

Results

Induction of diabetes

Induction of chronic high hyperglycaemia after injection of STZ (60 mg kg⁻¹, i.p.) was confirmed in rats by repeatedly measuring of fasting blood glucose levels above 17 mmol L⁻¹. During the t-MCAO procedure, fasting glucose levels in rats were measured before, immediately and 24 hours after t-MCAO. Fasting glucose levels in diabetic rats measured before and immediately after t-MCAO ranged from 22.6–31.2 mmol L⁻¹, whereas fasting blood glucose levels measured 24 hours after t-MCAO ranged from 17–33 mmol L⁻¹. In control non-diabetic animals fasting blood glucose levels measured at indicated time points were 5–9 mmol L⁻¹. Rat body weight was similar in all experimental groups (220–270 g).

Induction of t-MCAO

Successful t-MCAO was performed on 30 rats of a total of 36 rats used in this study. The remaining six animals were excluded from this study according to the exclusion criteria (see Methods section). Two diabetic rats in the 90-minute ischaemia group died within 24 hours after t-MCAO due to development of massive stroke. Furthermore, two animals in the 60-minute t-MCAO group were excluded from the study, one animal died of a large stroke and the other one was excluded due to subarachnoid haemorrhage, as confirmed at post-mortem examination of the brain. Two rats were excluded due to failure to reduce intra-ischaemic rCBF higher than 65% of baseline values, one diabetic animal in the 45-minute t-MCAO group and the other one in the control non-diabetic group.

Using LDF, rCBF was continuously monitored to confirm adequate reduction of intra-ischaemic rCBF (above 65% of baseline value) and reperfusion after removal of the occluding filament. Average intra-ischaemic reduction of rCBF was similar in all experimental groups, diabetic and non-diabetic, and ranged from 68–75% of baseline values (Figure 1(a)). rCBF monitored during the first 20-minutes of reperfusion did not significantly differ among the 30-minute t-MCAO control non-diabetic and 20, 30, 45 and 60-minute t-MCAO diabetic groups, ranging from 80–125% of baseline values (Figure 1(b)). Reperfusion was significantly impaired in all diabetic animals in the 90-minute t-MCAO group compared to all other groups and rCBF during reperfusion ranged from 56–62% of baseline values (Figure 1(b)).

Brain infarct volume in various durations of t-MCAO protocols

The volume of infarct was quantitatively measured by TTC staining 24 hours after t-MCAO. In the control group of rats without diabetes, 30-minutes of t-MCAO did not induce infarct of brain tissue on TTC staining, indicating that an ischaemia duration of 30-minutes is not sufficient to produce stroke in healthy animals (Table I, Figures 2 and 3).

In rats with diabetes, there was a significant increase in brain infarct volume expressed as a percentage of ischaemic

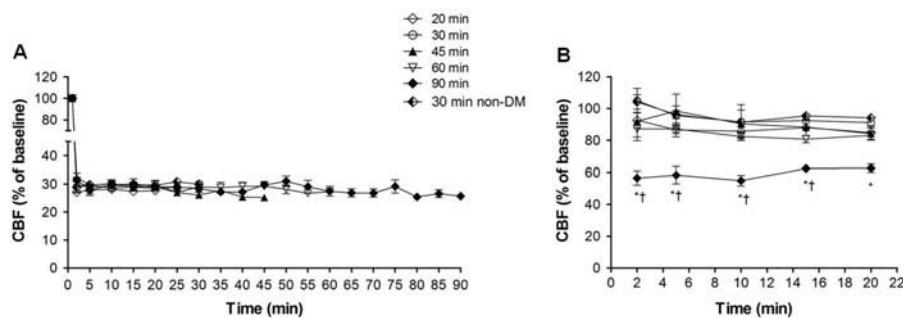


Figure 1. Cerebral blood flow in study groups during various durations of transitory middle cerebral artery occlusion and subsequent reperfusion period. Presents cerebral blood flow (CBF) during various durations (20, 30, 45, 60 and 90 minutes) of transitory middle cerebral artery occlusion (t-MCAO, a) and subsequent reperfusion period (a). (a) Similar reduction in CBF in all tested groups, proving successful t-MCAO in all study groups. Ninety minutes of t-MCAO resulted in significantly impaired reperfusion compared to other groups (b). * $p < 0.05$ compared to 20, 30 and 45 minute t-MCAO in diabetic (DM) rats and 30 minute t-MCAO in healthy control (non-DM) rats, † $p < 0.05$ compared to 60 minute t-MCAO in DM rats (two-way ANOVA RM and Bonferroni post-hoc test).

Table I. Proportion of the cortical, sub-cortical and total infarct in the overall hemisphere volume after various durations of transitory middle cerebral artery occlusion. The table shows the proportion of the sub-cortical, cortical and total infarct in the overall hemisphere volume, after 20, 30, 45, 60 and 90 minutes of transitory middle cerebral artery occlusion (t-MCAO, $n = 5/\text{group}$) in diabetic (DM) rats and 30 minutes t-MCAO in healthy control (non-DM) rat. Twenty minutes of t-MCAO was not successful in inducing brain infarct or resulted in an infarct of minimal size. Thirty, 45 and 60 minute t-MCAO resulted in a significantly larger infarct in DM rats. There was no significant difference in overall size of the infarct depending on the duration of the t-MCAO from 30–60 minutes; however, 90 minutes of t-MCAO resulted in an infarct that affected almost 90% of the total hemisphere volume. In non-DM, 30 minutes t-MCAO did not produce brain infarct.

t-MCAO duration	Non-diabetic rats		Diabetic rats			
	30 minutes	20 minutes	30 minutes	45 minutes	60 minutes	90 minutes
Total infarct	0%	$3 \pm 5\%$	$46 \pm 7\%$	$52 \pm 5\%$	$59 \pm 3\%$	$89 \pm 6\%$
Cortical infarct	0%	$1.5 \pm 2\%$	$34 \pm 7\%$	$35 \pm 10\%$	$40 \pm 5\%$	$67 \pm 6\%$
Striatal infarct	0%	$2 \pm 3\%$	$12 \pm 2\%$	$17 \pm 2\%$	$19 \pm 3\%$	$22 \pm 3\%$

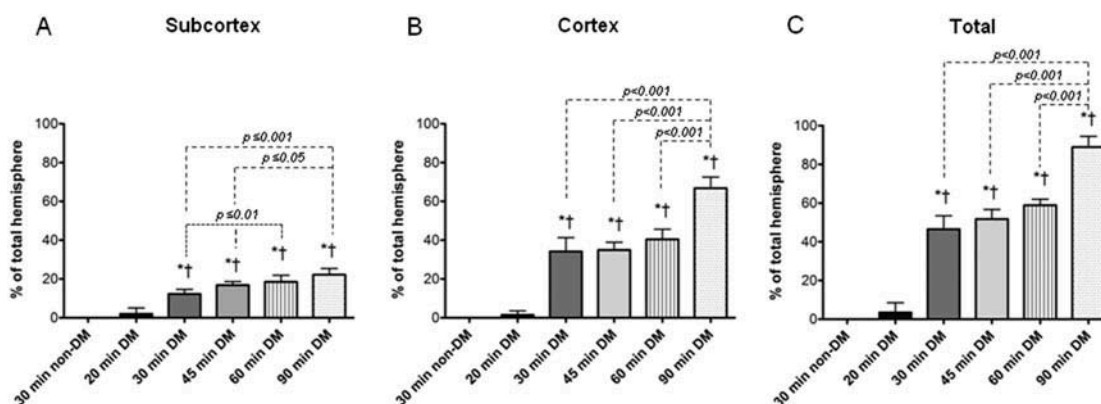


Figure 2. Proportion of the cortical, sub-cortical and total infarct in the total hemisphere volume after various duration of transitory middle cerebral artery occlusion. Shows the proportion of the sub-cortical (a), cortical (b) and total infarct in the overall hemisphere volume (c) after 20, 30, 45, 60 and 90 minutes of transitory middle cerebral artery occlusion (t-MCAO, $n = 5/\text{group}$) in diabetic (DM) rats and 30 minute t-MCAO in healthy control (non-DM) rats. While 20-minutes of t-MCAO was not successful in inducing brain infarct or resulted in infarct of minimal size; 30, 45 and 60 minute t-MCAO resulted in a significantly larger infarct in DM rats. There was no significant difference in overall size of infarct depending on duration of the t-MCAO from 30–60 minutes; however, 90 minutes of t-MCAO affected almost 90% of the total hemisphere volume (c). In non-DM rats there was no evidence of brain infarct. Data are presented as mean \pm SD; * $p < 0.05$ compared to 20 minute t-MCAO in DM rats; † $p < 0.05$ compared to 30 minute t-MCAO in non-DM rats; other relations are denoted at the panels (one-way ANOVA and Tukey post-hoc test).

hemispheres, among the experimental groups with longer duration of ischaemia (Table I, Figures 2 and 3).

Twenty minutes of t-MCAO in diabetic rats resulted in a significantly smaller infarct volume compared to all other durations of t-MCAO (Figures 2(a–c) and 3), producing only $3 \pm 5\%$ of an ischaemic hemisphere. The affected area was mostly striatal and only a small part was cortical infarct ($2 \pm 3\%$ vs $1.5 \pm 2\%$ of an ischaemic hemisphere, respectively; Table I, Figures 2 and 3). Thirty minutes of t-MCAO produced a significantly larger infarct that extended over $46 \pm 7\%$ of the ischaemic hemisphere, encompassing $12 \pm 2\%$ of striatum and $34 \pm 7\%$ of cortical area, which represents approximately half of the whole striatal area and half of the ipsilateral MCA irrigated cortex area (Figures 2(a–c) and 3, Table I).

Increasing ischaemia to 45 and 60 minutes resulted in spreading of infarct to almost the whole striatum that covers 22% of the hemisphere ($17 \pm 2\%$ in the 45-minute t-MCAO and $19 \pm 3\%$ in the 60-minute t-MCAO groups) and a large portion of the cortex ($35 \pm 10\%$ in the 45-minute t-MCAO and $40 \pm 5\%$ in the 60-minute t-MCAO group; cortical area irrigated by MCA is $\sim 66\%$ of the hemisphere). Total infarct size was $52 \pm 5\%$ in the 45-minute t-MCAO and $59 \pm 3\%$ of

the ischaemic hemisphere in the 60-minute t-MCAO (Table I, Figures 2(a–c) and 3).

The longest ischaemia duration of 90-minutes in diabetic rats led to massive hemispheric stroke ($89 \pm 6\%$ of hemisphere; the whole striatum ($22 \pm 3\%$) and the whole MCA territory ($67 \pm 6\%$)) (Table I, Figures 2(a–c) and 3).

Discussion

The most important findings in the present study are: (a) diabetic female SD rats are highly sensitive to ischaemia in a t-MCAO stroke model; (b) 60 and 90-minute t-MCAO currently used in non-diabetic female rats seems to be too long for diabetic ones, producing massive strokes in rats with diabetes; (c) lesser reperfusion was found in longer ischaemia in the 90-minute group (which possibly could be explained as a result of already irreversible brain infarct with brain vascular derangement). This brain vascular sign could be a marker of point of no return in stroke treatment; (d) 30-minute t-MCAO seems to be an adequate duration of occlusion for the diabetic rat female stroke model, consistently producing medium sized stroke which affects 30–50% of ischaemic

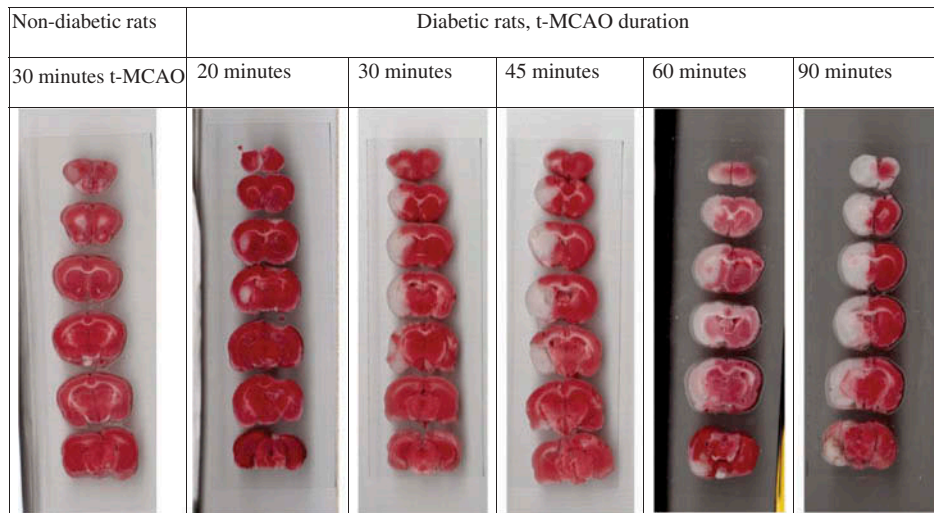


Figure 3. Brain infarct in the control group and after various duration of transitory middle cerebral artery occlusion in experimental groups. Typical brain infarct in control group and after 20, 30, 45, 60 and 90 minutes of transitory middle cerebral artery occlusion in experimental groups (t-MCAO, $n = 5/\text{group}$) in diabetic (DM) rats and 30 minutes t-MCAO in healthy control (non-DM) rats (t-MCAO, $n = 5/\text{group}$). While 30 minutes t-MCAO in healthy controls did not produce cerebral infarction and 20 minutes of t-MCAO was not successful in inducing brain infarction or resulted in infarction of minimal size; 30, 45 and 60 minutes of t-MCAO resulted in significantly larger infarction in DM rats.

hemisphere; (e) 20-minute t-MCAO in rats with diabetes produced small striatal strokes of inconsistent size; and (f) it is questionable to compare results of occlusion for rats with and without diabetes, even if the duration of t-MCAO is equal.

Sprague-Dawley rat strain (SD) is the most widely used strain in rat MCAO experiments (41% of researchers have done experiments on this strain, vs 34% on Wistar, 10% on SHR, 2% on Long-Evans and 2% other strains [30]). SD rats have lower complication rates including subarachnoid haemorrhage (SAH) compared to Fisher and Wistar rats, due to anatomical course of ICA in petrous segment [30]. Previous experiments suggested that oestrogen relatively protected female rats from diabetes-exacerbated stroke damage [18]. However, present experiments showed that the diabetic female rats had extremely high susceptibility to brain infarction. When establishing a model for stroke in diabetic rats based on a published model in rats without diabetes, at first it was found that the usual duration of ischaemia from 60–120 minutes has produced large strokes, with massive oedema and devastating neurological deficits, such as inability to feed, immobility and alteration of consciousness. These strokes were very often lethal in the first 24 hours after the experiment, mostly due to massive oedema and a rise in intracranial pressure. Similarly, patients with the most severe strokes of the whole MCA territory and high NIHSS (National Institute of Health Stroke Score) are poor candidates for treatment with thrombolysis and mostly die due to brain oedema and complications of dysphagia and immobility, but also have higher risk of secondary haemorrhage. Having this in mind, in the present experiments, duration of MCA occlusion was shortened variously in the groups to determine MCAO duration that will result in moderate stroke, already described in non-diabetic patients/rats, i.e. 30 and 50% of volume of hemisphere [31–34]. In the present experiments longer ischaemia inevitably produced a larger brain infarct (Figures 2(c) and 3, Table I) and usual duration of ischaemia in rats without

diabetes (in earlier studies mostly 90 minutes, but also 60–120 minutes) was too long for diabetic ones [7]. Ischaemia of 60 and 90 minutes produced massive infarct with lesser probability for survival as well as any kind of treatment.

Diabetes worsens stroke outcome; diabetic patients have 2–4-times higher risk for stroke compared to non-diabetic patients [13]. Possible underlying causes are chronic hyperglycaemia, which leads to free oxygen radicals and cytokines production and increases ischaemic brain cells pre-disposition to apoptosis [35]. In addition, the intimal artery thickening and arteriolar occlusion occurs in diabetes, contributing by impaired vascular function to inadequate tissue perfusion [12]. In the past research, animal models of longer ischaemia and longer time before treatments of stroke starts were accompanied with negative or poor results in outcome of treatments [31]. For example, rTPA treatment after 2 hours of t-MCAO showed no effect on brain infarct size and neurological deficit, but had negative effects on the risk of intracerebral haemorrhage, blood–brain barrier breakdown and production of inflammatory mediators [10]. Longer ischaemia inevitably worsens brain infarct and augments inflammatory response that is related to vascular disease and diabetes [36]. Higher inflammatory response leads to brain oedema and consequently to lower brain perfusion and incurable damage [37]. Taking into account all these facts, a diabetic rat model of stroke should be modified to suit the purpose, i.e. ischaemia duration needs to be shortened to produce a medium-sized brain infarct that would mimic treatable human stroke. Otherwise, the false conclusion can be drawn from massive strokes due to an inadequate experimental design—that no therapy is useful in stroke treatment in diabetics. In clinical practice, the majority of strokes are effectively treated in a short period after they occur and, in fact, they are of medium and smaller size. On the other side, a longer duration of ischaemia and, consequently, larger strokes are mostly resistant to treatment and detrimental to patients.

In the present study, a 20-minute ischaemia duration did not produce infarction in two out of five rats with diabetes, while in the remaining animals the infarct was rather small ($3 \pm 5\%$ of ischaemic hemisphere) and located almost only in the striatum (Table I, Figures 2(a) and (b) and 3).

Only 30-minutes of ischaemia in rats with diabetes consistently produced medium sized infarcts that include $\sim 50\%$ of the cortex and $\sim 50\%$ of striatum irrigated by MCA (Figures 2(a) and (b) and 3, Table I). That corresponds well with large artery atherosclerotic stroke [38]. At the same time, 30 minutes of occlusion was not devastating or lethal for rats, making it an optimal duration of ischaemia in a stroke model for future research. Brain infarcts of the same size were used successfully in rats without diabetes [8,31].

Longer ischaemia durations of 45 and 60 minutes in rats with diabetes produced larger stroke in the MCA territory, encompassing almost the whole striatum and a large portion of the cortex (Table I, Figures 2(a) and (b) and 3). Ischaemia of 90 minutes led to massive hemispheric stroke (Table I and Figures 2(b) and 3). Such large strokes would be in clinical practice almost always incompatible with survival due to development of brain oedema, raised intracranial pressure (ICP) with secondary low arterial perfusion and subsequent spreading of infarction, brain herniation, etc. This type of stroke occurs after sudden and permanent MCA occlusion, with no reperfusion and without preformed arterial collaterals. One significant finding of this study was also lesser reperfusion in longer ischaemia. All animals in the 90-minute group had significantly lower reperfusion (56–62% of baseline compared to 80–125% of baseline in other groups, Figure 1(b)) and brain vascular derangement in which normal post-ischaemic reperfusion as a salvation mechanism does not appear. It is speculated that this brain vascular sign is important and could be a marker of point of no return in stroke treatment.

If compared to humans, strokes classified of small size have a range between 4.5–14% of the ipsilateral hemisphere [39,40]. Infarct is considered malignant when it covers more than 50% of the cerebral hemisphere, but some studies report that malignant infarction occurs when the stroke size is greater than 39% of the hemisphere [41,42].

For every experimental model that could represent human conditions, it is important to simulate average situation. The model has to simulate most common types of stroke in humans which are treatable strokes—those of medium and smaller size. Based on the results of the present study, the best model for mimicking human condition of stroke would be 30-minute occlusion that led to infarct of approximately half of the striatum and half of the cortex irrigated by MCA. All other longer models produced total or almost total striatum devastation and the cortex was almost totally destroyed in the 90-minute model, which is incurable in human medicine and there is no purpose to further explore them.

Conclusion

The present study tends to impact the experimental design of stroke model in basic research and consequently influence clinical research in the field of cerebrovascular disorders. It

is proposed that duration of ischaemia in a female rat model has to be adjusted depending on which type of stroke one wants to produce. Not all strokes are treatable the same way and not all of them have the same prognosis; smaller and medium size strokes have the best chances for improvement by therapy. Almost any treatment of massive stroke or treatments started after certain time points after stroke tend to be ineffective. The present study, although restricted to female SD rats, aims to alter the current point of view on duration of t-MCAO used in the experiments. The main application of the results of the present study are in finding the optimal duration of MCA occlusion in rats with diabetes in stroke model that could be effectively treated. In addition, by establishing the optimal duration of occlusion, results of different studies would be comparable and testing of new therapeutic options in rat stroke experiments would be easier and more effective, finally leading to better understanding of mechanisms of prevention of neurological deficits. In the present study, the 30-minute t-MCAO model in female rats with diabetes seems to produce a stroke of medium size, most likely the best for the experimental and clinical research on new treatment options.

Declaration of interest

The authors report no conflicts of interest. Support came from a grant by the Ministry of Science, Education and Sport of Republic of Croatia #219-2160133-2034 (principal investigator: Ines Drenjančević) and by EU FP7 grant GlowBrain (REGPOT-2012-CT2012-316120).

References

- Williams LS, Rotich J, Qi R, Fineberg N, Espay A, Bruno A, Fineberg SE, Tierney WR. Effects of admission hyperglycemia on mortality and costs in acute ischemic stroke. *Neurology* 2002;59:67–71.
- Kaarisalo MM, Raiha I, Sivenius J, Immonen-Raiha P, Lehtonen A, Sarti C, Mähönen M, Torppa J, Tuomilehto J, Salomaa V. Diabetes worsens the outcome of acute ischemic stroke. *Diabetes Research & Clinical Practice* 2005;69:293–298.
- Janghorbani M, Hu FB, Willett WC, Li TY, Manson JE, Logroscino G, Rexrode KM. Prospective study of type 1 and type 2 diabetes and risk of stroke subtypes: the Nurses' Health Study. *Diabetes Care* 2007;30:1730–1735.
- Soedamah-Muthu SS, Fuller JH, Mulnier HE, Raleigh VS, Lawrenson RA, Colhoun HM. High risk of cardiovascular disease in patients with type 1 diabetes in the U.K.: a cohort study using the general practice research database. *Diabetes Care* 2006;29:798–804.
- Secrest AM, Prince CT, Costacou T, Miller RG, Orchard TJ. Predictors of and survival after incident stroke in type 1 diabetes. *Diabetes & Vascular Disease Research* 2013;1:3–10.
- Jauch EC, Saver JL, Adams HP Jr., Bruno A, Connors JJ, Demaerschalk BM, Khatri P, McMullan PW Jr., Qureshi AI, Rosenfield K, et al. Guidelines for the early management of patients with acute ischemic stroke: a guideline for healthcare professionals from the American Heart Association/American Stroke Association. *Stroke* 2013;44:870–947.
- Nagasawa H, Kogure K. Correlation between cerebral blood flow and histologic changes in a new rat model of middle cerebral artery occlusion. *Stroke* 1989;20:1037–1043.
- Lan X, Qu H, Yao W, Zhang C. Granulocyte-colony stimulating factor inhibits neuronal apoptosis in rat model of diabetic cerebral ischemia. *Tohoku Journal of Experimental Medicine* 2008;216:117–126.

9. Fan X, Qiu J, Yu Z, Dai H, Singhal AB, Lo EH, Wang X. A rat model of studying tissue-type plasminogen activator thrombolysis in ischemic stroke with diabetes. *Stroke* 2012;43:567–570.
10. Ning R, Chopp M, Yan T, Zacharek A, Zhang C, Roberts C, Cui X, Lu M, Chen J. Tissue plasminogen activator treatment of stroke in type-1 diabetes rats. *Neuroscience* 2012;222:326–332.
11. Elgebaly MM, Oghi S, Li W, Mezzetti EM, Prakash R, Johnson MH, Bruno A, Fagan SC, Ergul A. Neurovascular injury in acute hyperglycemia and diabetes: a comparative analysis in experimental stroke. *Translational Stroke Research* 2011;2:391–398.
12. Chen J, Ye X, Yan T, Zhang C, Yang XP, Cui X, Cui Y, Zacharek A, Roberts C, Liu X, et al. Adverse effects of bone marrow stromal cell treatment of stroke in diabetic rats. *Stroke* 2011;42:3551–3558.
13. Kusaka I, Kusaka G, Zhou C, Ishikawa M, Nanda A, Granger DN, Zhang JH, Tang J. Role of AT1 receptors and NAD(P)H oxidase in diabetes-aggravated ischemic brain injury. *American Journal of Physiology Heart & Circulatory Physiology* 2004;286:2442–2451.
14. Ye X, Yan T, Chopp M, Zacharek A, Ning R, Venkat P, Roberts C, Chen J. BMSC and Niaspan treatment of stroke enhances white matter remodeling and synaptic protein expression in diabetic rats. *International Journal of Molecular Sciences* 2013;14:22221–22232.
15. Ye X, Chopp M, Liu X, Zacharek A, Cui X, Yan T, Roberts C, Chen J. Niaspan reduces high-mobility group box 1/receptor for advanced glycation end products after stroke in type-1 diabetic rats. *Neuroscience* 2011;190:339–345.
16. Ye X, Chopp M, Cui X, Zacharek A, Cui Y, Yan T, Shehadah A, Roberts C, Liu X, Lu M, et al. Niaspan enhances vascular remodeling after stroke in type 1 diabetic rats. *Experimental Neurology* 2011;232:299–308.
17. Simard JM, Yurovsky V, Tsybalyuk N, Melnichenko L, Ivanova S, Gerzanich V. Protective effect of delayed treatment with low-dose glibenclamide in three models of ischemic stroke. *Stroke* 2009;40:604–609.
18. Toung TK, Hurn PD, Traystman RJ, Sieber FE. Estrogen decreases infarct size after temporary focal ischemia in a genetic model of type 1 diabetes mellitus. *Stroke* 2000;31:2701–2706.
19. Niessen F, Hilger T, Hoehn M, Hossmann KA. Differences in clot preparation determine outcome of recombinant tissue plasminogen activator treatment in experimental thromboembolic stroke. *Stroke* 2003;34:2019–2024.
20. Wang CX, Yang T, Shuaib A. An improved version of embolic model of brain ischemic injury in the rat. *Journal of Neuroscience Methods* 2001;109:147–151.
21. Kong Q, Hafeez A, Yu W, Ren C, Geng X, Xiao Y, Liu S, Zhang Y, Mao R, Zhou J, Ding Y, Ji X. Acute recanalization of carotid stenosis is not proper: an experimental ischaemic stroke study. *Neurological Research* 2015;37:397–402.
22. Park SI, Park SK, Jang KS, Han YM, Kim CH, Oh SJ. Preischemic neuroprotective effect of minocycline and sodium ozagrel on transient cerebral ischemic rat model. *Brain Research* 2015;1599:85–92.
23. Akbarzadeh A, Norouzzian D, Mehrabi MR, Jamshidi Sh, Farhangi A, Verdi AA, Mofidian SM, Rad BL. Induction of diabetes by Streptozotocin in rats. *Indian Journal of Clinical Biochemistry* 2007;22:60–64.
24. Srinivasan K, Ramarao P. Animal models in type 2 diabetes research: an overview. *Indian Journal of Medical Research* 2007;125:451–472.
25. Koizumi J, Yoshida Y, Nakazawa T, Ooneda G. Experimental studies of ischemic brain edema. I.: a new experimental model of cerebral embolism in rats in which recirculation can be introduced in the ischemic area. *Japanese Journal of Stroke* 1986;8:1–8.
26. Zea Longa EL, Weinstein PR, Carlson S, Cummins R. Reversible middle cerebral artery occlusion without craniectomy in rats. *Stroke* 1989;20:84–91.
27. Zhang PB, Liu Y. Effects of ketamine-midazolam anesthesia on focal cerebral ischemic injury in rats. *Journal of First Military Medicine University* 2004;12:1337–1341.
28. Renic M, Klaus JA, Omura T, Kawashima N, Onishi M, Miyata N, Koehler RC, Harder DR, Roman RJ. Effect of 20-HETE inhibition on infarct volume and cerebral blood flow after transient middle cerebral artery occlusion. *Journal of Cerebral Blood Flow & Metabolism* 2009;29:629–639.
29. Bederson JB, Pitts LH, Germano SM, Nishimura MC, Davis RL, Bartkowski HM. Evaluation of 2,3,5-triphenyltetrazolium chloride as a stain for detection and quantification of experimental cerebral infarction in rats. *Stroke* 1986;17:1304–1308.
30. Dittmar MS, Vatankhah B, Fehm NP, Schuierer G, Bogdahn U, Horn M, Schlachetzki F. Fisher-344 rats are unsuitable for MCAO filament model due to their cerebrovascular anatomy. *Journal of Neuroscience Methods* 2006;156:50–54.
31. Carmichael ST. Rodent models of focal stroke: size, mechanism, and purpose. *NeuroRx* 2005;3:396–409.
32. Toung TK, Hurn PD, Traystman RJ, Sieber FE. Estrogen decreases infarct size after temporary focal ischemia in a genetic model of type 1 diabetes mellitus. *Stroke* 2000;31:2701–2706.
33. Wang Q, Peng Y, Chen S, Gou X, Hu B, Du J, Lu Y, Xiong L. Pretreatment with electroacupuncture induces rapid tolerance to focal cerebral ischemia through regulation of endocannabinoid system. *Stroke* 2009;40:2157–2164.
34. Dunn KM, Renic M, Flasch AK, Harder DR, Falck J, Roman RJ. Elevated production of 20-HETE in the cerebral vasculature contributes to severity of ischemic stroke and oxidative stress in spontaneously hypertensive rats. *American Journal of Physiology Heart & Circulatory Physiology* 2008;295:2455–2465.
35. Tsuruta R, Fujita M, Ono T, Koda Y, Koga Y, Yamamoto T, Nanba M, Shitara M, Kasaoka S, Maruyama I, et al. Hyperglycemia enhances excessive superoxide anion radical generation, oxidative stress, early inflammation, and endothelial injury in forebrain ischemia/reperfusion rats. *Brain Research* 2010;1309:155–163.
36. Schmidt AM, Yan SD, Yan SF, Stern DM. The multiligand receptor RAGE as a progression factor amplifying immune and inflammatory responses. *Journal of Clinical Investigation* 2001;108:949–955.
37. Melton JE, Nattie EE. Intracranial volume adjustments and cerebrospinal fluid pressure in the osmotically swollen rat brain. *American Journal of Physiology* 1984;246:533–541.
38. Adams HP Jr., Bendixen BH, Kappelle LJ, Biller J, Love BB, Gordon DL, Marsh EE 3rd. Classification of subtype of acute ischemic stroke. Definitions for use in a multicenter clinical trial: TOAST. *Trial of Org 10172 in Acute Stroke Treatment*. *Stroke* 1993;24:35–41.
39. Brott T, Marler JR, Olinger CP, Adams HP Jr., Tomsick T, Barsan WG, Biller J, Eberle R, Hertzberg V, Walker M. Measurements of acute cerebral infarction: lesion size by computed tomography. *Stroke* 1989;20:871–875.
40. Nopoulos P, Flaum M, O'Leary D, Andreasen NC. Sexual dimorphism in the human brain: evaluation of tissue volume, tissue composition and surface anatomy using magnetic resonance imaging. *Psychiatry Research* 2000;98:1–13.
41. Oppenheim C, Samson Y, Manai R, Lalam T, Vandamme X, Crozier S, Srouf A, Cornu P, Dormont D, Rancurel G, et al. Prediction of malignant middle cerebral artery infarction by diffusion-weighted imaging. *Stroke* 2000;31:2175–2181.
42. Mori K, Aoki A, Yamamoto T, Horinaka N, Maeda M. Aggressive decompressive surgery in patients with massive hemispheric embolic cerebral infarction associated with severe brain swelling. *Acta Neurochirurgica* 2001;143:483–491.